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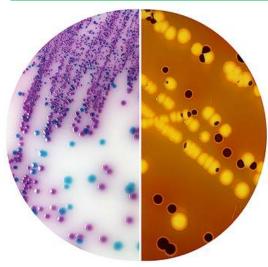


Biolife

INSTRUCTIONS FOR USE

BIOSECTOR CHROMOGENIC SALMONELLA AGAR XLD AGAR

Ready-to-use bi-plates



Salmonella sp. (magenta-red on CSA and black on XLD) and E.aerogenes (green on CSA and yellow on XLD.

1-INTENDED USE

In vitro diagnostic device. Selective and differential media for the isolation of Gram-negative enteric pathogens, especially Salmonella and Shigella, from clinical and non clinical specimens.

2 - COMPOSITIONS - TYPICAL FORMULAS

CHROMOGENIC SALMONELLA AGAR*

Peptones	10.0 g	Emulsifying agents	11.4 g
Selective compounds	12.0 g	Opacifier	10.0 g
Cefsulodin	5.0 mg	Agar	15.0 g
Chromogenic mixture	0.9 g	Purified water	1000 mL
XLD AGAR*			
Xylose	3.50 g	Sodium desoxycholate	2.50 g
L-lysine	5.00 g	Sodium thiosulphate	6.80 g
Lactose	7.50 g	Ferric ammonium citrate	0.80 g
Sucrose	7.50 g	Phenol red	0.08 g
Sodium chloride	5.00 g	Agar	13.50 g
Yeast extract	3.00 g	Purified water	1000 mL

^{*}The formula may be adjusted and/or supplemented to meet the required performances criteria.

3 - PRINCIPLE OF THE METHOD AND EXPLANATION OF THE PROCEDURE

The bi-plate with Chromogenic Salmonella Agar and XLD Agar culture media is indicated in clinical and industrial microbiology when determination in Salmonella spp. (including S.typhi and Salmonella lac + strains) and Shigella spp is required.

CHROMOGENIC SALMONELLA AGAR

Chromogenic Salmonella Agar is a selective and diagnostic medium useful for the isolation of Salmonella spp. from clinical and non clinical specimens and for the presumptive identification of the colonies. Chromogenic Salmonella Agar is included by ISTISAN Report¹ in the plating media range for the detection of Salmonella spp. and chromogenic media are included as the second plating medium in ISO Standards for detection of Salmonella in food and water. 2,3

Peptones provide carbon, nitrogen, vitamins and trace elements for bacterial growth. The selective compounds incorporated in the medium are the following: cefsulodin, a third generation cephalosporin antibiotic that has very specific activity against P. aeruginosa and S.aureus, sodium desoxycholate that suppresses the growth of Gram-positive and some Gram-negative bacteria and Tergitol 4, active mainly against the growth of *Proteus* spp.

Differentiation of Salmonella from the other organisms that grow is achieved by:

- a chromogenic substrate for C₈ esterase enzyme, that is split by Salmonella spp. with the release of an insoluble magenta-red
- a chromogenic glucopyranoside derivative which is split by β -glucosidase with the release of an insoluble blue-green chromophore.

Some Enterobacteriaceae, including Klebsiella and Enterobacter, but not Salmonella, are ß-glucosidase positive and if growing will form blue-green or dark blue colonies, even if they are esterase positive, which make them easy to differentiate from magenta-red Salmonella colonies. The chromogenic and selective compounds of the medium also allow the detection of the rare lactose positive Salmonella strains, missed on traditional media based on lactose fermentation. Chromogenic Salmonella Agar is useful also for the detection of S.Typhi and S.Paratyphi. The ready-to-use plates include a compound that gives a white opaque background to the medium and enhances the colour of the colonies.

XLD AGAR

XLD Agar is a selective and differential medium intended for the isolation of Gram-negative enteric pathogens, especially Salmonella and Shigella from clinical specimens. 46 It is recommended for the detection of Salmonella in non sterile pharmaceutical products according to harmonized EP, USP, JP method⁷ and by FDA-BAM for detection of *Salmonella* in food⁸.

Yeast extract provides carbon, nitrogen, vitamins and trace elements for bacterial growth; sodium chloride maintains the osmotic balance in the medium; sodium desoxycholate is a selective agent for suppressing the growth of Gram positive bacteria. XLD Agar contains three indicator systems: xylose, lactose, and sucrose combined with phenol red, lysine hydrochloride and again phenol red, sodium thiosulfate and ferric ammonium citrate. Target bacteria are tentatively grouped by reading the effect of carbohydrate fermentation, lysine decarboxylation and formation of hydrogen sulphide.

4 - PHYSICAL CHARACTERISTICS

CHROMOGENIC SALMONELLA AGAR Medium appearance

Final pH at 20-25 °C XLD AGAR

Medium appearance Final pH at 20-25 °C whitish opaque

 7.2 ± 0.2

red, limpid 7.4 ± 0.2







5 - MATERIALS PROVIDED - PACKAGING

Product	Туре	REF	Pack
Biosector Chromogenic Salmonella Agar / XLD Agar	Ready-to-use bi-plates	495351	2 x 10 plates ø 90 mm with 2 sectors primary packaging: 2 cellophane sachets secondary packaging: cardboard box

6 - MATERIALS REQUIRED BUT NOT PROVIDED

Sterile loops and swabs, incubator and laboratory equipment as required, ancillary culture media and reagents for the identification of the colonies.

7 - SPECIMENS

Biosector Chromogenic Salmonella Agar / XLD Agar is intended for the bacteriological processing of clinical specimens such as faeces, rectal swab, urine, bile, 4-6 and non clinical specimens such as non-sterile pharmaceutical products and food 2,3,8. Collect specimens before antimicrobial therapy where possible. Good laboratory practices for collection, transport and storage of clinical specimens should be applied.⁹ Consult appropriate standard methods for details of collection and preparation of non-clinical specimens.^{2,3,7,8}

8 - TEST PROCEDURE

Allow plates to come to room temperature and to dry the surface of the medium.

Inoculate and streak the specimen with a loop over the media surfaces to obtain well isolated colonies. Alternatively, if the material is being cultured directly from a swab, roll the swab over a small area of the surface at the edge; then streak from this inoculated area. Maximal recovery of Salmonella from faecal specimens is obtained by using an enrichment step in Selenite Broth. For Shigella isolation from faecal specimens, the enrichment in GN Broth is advised.6

Incubate inoculated bi-plates with the specimen or with a specimen enriched in liquid medium, in aerobic conditions at 35-37°C for 18-24 hours. Consult appropriate references for the detection of Shigella and Salmonella in non clinical specimens. 2,3,7

9 - READING AND INTERPRETATION

After incubation, observe the bacterial growth and record the specific morphological and chromatic characteristics of isolated colonies. The different microorganisms grow on Chromogenic Salmonella Agar with the following characteristics:

Microorganism	Growth characteristics
Salmonella spp.	good growth, magenta-red colonies
Salmonella spp. lac+	good growth, magenta-red colonies
Salmonella Typhi	good growth, magenta-red colonies
E.coli	poor growth with colourless colonies
Enterobacter spp.	growth with blue-green colonies
Klebsiella spp.	poor growth with blue-green colonies
Pseudomonas spp.	inhibited
Proteus spp.	poor growth with pale brown or green colonies
Gram-positive bacteria	inhibited

Interpretation of colonies' colours on XLD Agar¹⁰:

Red colonies: alkaline reaction, non-fermentation of xylose/sucrose/lactose, or fermentation of xylose followed by decarboxylation of lysine: possible Shigella or Providencia or Pseudomonas spp. or Salmonella sp. H₂S negative

Red colonies with black centre: xylose fermentation only, lysine positive, H₂S positive, rapid depletion of xylose and resultant alkalinity due to lysine decarboxylation, black centre due to H2S production possible only in alkaline pH environment: suspect Salmonella H2S

Opaque yellow colonies: xylose fermentation, lysine negative and non fermentation of lactose and sucrose, acid pH: possible E.coli, Klebsiella/Enterobacter, Citrobacter, Serratia, Proteus spp.

Yellow colonies: lactose or sucrose fermentation, lysine negative, acid pH: possible coliforms or sucrose-positive P. vulgaris

10 - USER QUALITY CONTROL

All manufactured lots of the product are released for sale after the Quality Control has been performed to check the compliance with the specifications. However, it is responsibility of the end-user to perform Quality Control testing in accordance with the local applicable regulations, in compliance with accreditation requirements and the experience of the Laboratory. Here below are listed some test strains useful for the quality control.11

CONTROL STRAINS	INCUBATION T°/T/ATM	EXPECTED RESULTS
CHROMOGENIC SALMONELLA AGAR		
S.Typhimurium ATCC 14028	35-37°C / 18-24h / A	growth, magenta-red colonies
S. Enteritidis ATCC 13076	35-37°C / 18-24h / A	growth, magenta-red colonies
E. aerogenes ATCC 13048	35-37°C / 18-24h / A	growth, blue-green colonies
P. aeruginosa ATCC 27853	35-37°C / 18-24h / A	inhibited
XLD Agar		
S.Typhimurium ATCC 14028	30-35 or 35-37°C / 18-24h / A	growth, red colonies with black centre
S.flexneri ATCC 12022	30-35 or 35-37°C / 18-24h / A	growth, red colonies
E.faecalis ATCC 29212	30-35 or 35-37°C / 18-24h / A	inhibited
E.coli ATCC 25922	30-35 or 35-37°C / 18-24h / A	partially inhibited, yellow colonies

A: aerobic incubation; ATCC is a trademark of American Type Culture Collection Incubation temperature depends of the followed Standard (CLSI¹³ or EuPh⁷)



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11 - PERFORMANCES CHARACTERISTICS

Prior to release for sale, representative samples of all lots of ready to use bi-plates of Chromogenic Salmonella Agar / XLD Agar and of the raw materials used for the production of bi-plates (dehydrated Chromogenic Salmonella Agar REF 405350 supplemented with Salmonella Selective Supplement REF 4240013 and XLD Agar, REF 402206) are tested for productivity and selectivity by comparing the results with a previously approved Reference Batch.

CHROMOGENIC SALMONELLA AGAR

Productivity is tested by a quantitative test with the target strains S.Enteritidis ATCC 13076 and S.Typhimurium ATCC 14028; Chromogenic Salmonella Agar plates are inoculated with decimal dilutions in saline of a suspension of colonies and incubated at 35-37°C for 18-24 hours. The colonies are enumerated on Test Batch (TB) and Reference Batch (RB) and the productivity ratio (Pr=UFC_{TB}/UFC_{RB}) is calculated. If Pr is ≥ 0.7 and if the colonies' colour is typical (magenta red colonies) the results are considered acceptable and conform to the specifications

Specificity is evaluated with by semi-quantitative ecometric technique with the following non-target strains: S.flexneri ATCC 12022, E.aerogenes ATCC 13048, E.coli ATCC 8739; E.aerogenes grows with blue-green colonies, the growth of S.flexneri is not inhibited and the colonies are colourless, *E.coli* is partially inhibited with colourless colonies. Selectivity is evaluated with modified Miles-Misra surface drop method by inoculating the plates with decimal dilutions in saline from 10⁻¹ to 10⁻⁴ of a 0.5 McFarland suspension of the non-target strains P.vulgaris ATCC 13315, A.calcoaceticus ATCC 19606, P.aeruginosa ATCC 27853, A.hydrophila ATCC 7966, E.faecalis ATCC 19433, Mucor CBM1. A.hydrophila is partially inhibited and grows with magenta colonies; the growth of other non-target strains is totally inhibited at the dilution 10-1

XLD AGAR

Productivity is tested by a quantitative test with 2 target strains: S. Enteritidis ATCC 13076, S.Typhimurium ATCC 14028; XLD Agar plates are inoculated with decimal dilutions in saline of the colonies' suspensions and incubated at 30-35°C for 18-24 hours. The colonies are enumerated on both batches and the productivity ratio (Pr) is calculated. If Pr is ≥ 0,7 and if the colonies morphology and colour are typical (red colonies with black centre) the results are considered acceptable and conform to the specifications. Furthermore the productivity characteristics are tested by semi-quantitative ecometric technique with the target strain S.flexneri ATCC 12022. After incubation, colonies' colour and the amount of growth is evaluated and recorded.

Selectivity is evaluated with modified Miles-Misra surface drop method by inoculating the plates with decimal dilutions in saline from 10⁻¹ to 10⁻³ of a 0.5 McFarland suspension of the non-target strains E.faecalis ATCC 19433 and E.coli ATCC 25922. The growth of non-target strain E.faecalis is inhibited at the dilution 10⁻¹, the growth of Gram negative non-target strain is partially inhibited and the colonies show typical yellow colour, according to the specifications.

Chromogenic Salmonella Agar was evaluated by Babic-Ergeg et al. 10 on 3,000 stool specimens, 45 of which positive for Salmonella, including SS Agar as the reference medium. The authors reported a sensitivity of 100% and a specificity of 99% in the isolation and preliminary identification of Salmonella colonies.

In another independent study¹¹, 50 pure cultures of Salmonella of clinical origin, gave all the specific chromatic reactions; among the other 80 strains of Gram negative bacteria tested, not belonging to the Salmonella genus, 3 out of 3 strains of P. aeruginosa and 1 out of 3 strain of A.baumannii provided chromatic results similar to Salmonella spp. (red-pink colonies), the remaining 76 strains of Enterobacteriaceae gave non-typical chromatic reactions; 20 out of 20 strains of Gram positive bacteria were totally inhibited.

Chromogenic Salmonella Agar performance was evaluated with an in-house study, compared to Hektoen Enteric Agar (HEA). Productivity, selectivity and specificity have been evaluated by semi-quantitative ecometric technique, incubating at 35-37°C for 18-24 hours, using 43 bacterial strains: 8 target strains and 35 non target strains. 8 Salmonella strains, including 2 S.Typhi, showed a good growth with magenta colonies; 3 Shigella strains showed a poorer growth than on HEA with colourless colonies; 22 Enterobacteriaceae strains belonging to 9 genera showed a poorer growth than on HEA with colourless or blue-green colonies; 4 P.aeruginosa strains were totally inhibited; 2 non fermenters strains were totally inhibited and A.hydrophila grew with magenta red colonies; 1 Gram positive strain was totally inhibited and 1 yeast strain was partially inhibited showing colourless colonies.

12 - LIMITATIONS OF THE METHOD

- Some strains of Pseudomonas, Acinetobacter and Aeromonas, resistant to antimicrobial agents of Chromogenic Salmonella Agar, may grow with red-pink colonies, differentiable from Salmonella with oxidase test.
- The growth rate on the plates also depends on the nutritional requirements of Salmonella. It is possible that some strains with particular metabolic characteristics may not grow on Chromogenic Salmonella Agar or grow colourless (e.g., Salmonella enterica serovar Dublin grows with white colonies).
- On XLD Agar non-enteric organisms such as Pseudomonas may grow; Pseudomonas and Providencia rettgeri may both exhibit red colonies. Some Proteus spp. may develop black centres. 10
- On XLD Agar S.Parathyphi A, S.Cholerae-suis, S.Pullorum and S.Gallinarum may form red colonies without black centre, thus resembling Shigella spp.
- Incubation exceeding 48 hours may lead to false positive results.¹⁰
- Even if the microbial colonies on the plates are differentiated on the basis of their morphological and chromatic characteristics, it is recommended that biochemical, immunological, molecular, or mass spectrometry testing be performed on isolates, from pure culture, for complete identification. If relevant, perform antimicrobial susceptibility testing.
- This culture medium is intended as an aid in the diagnosis of infectious disease; the interpretation of the results must be made considering the patient's clinical history, the origin of the sample and the results of other diagnostic tests.

13 - PRECAUTIONS AND WARNINGS

- · This product is a qualitative in vitro diagnostic, for professional use only; it is to be used by adequately trained and qualified laboratory personnel, observing approved biohazard precautions and aseptic techniques.
- This product is not classified as dangerous according to current European legislation.
- These culture media contain raw materials of animal origin. The ante and post mortem controls of the animals and those during the production and distribution cycle of the raw materials, cannot completely guarantee that these products do not contain any transmissible pathogen. Therefore, it is recommended that the ready-to use plates be treated as potentially infectious, and handled observing the usual specific precautions: do not ingest, inhale, or allow to come into contact with skin, eyes, mucous membranes. Download the TSE Statement from the website www.biolifeitaliana.it, describing the measures implemented by Biolife Italiana S.r.l. for the risk reduction linked to infectious animal diseases.
- All laboratory specimens should be considered infectious.
- The laboratory area must be controlled to avoid contaminants such as culture medium or microbial agents.

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- Each bi-plate is for single use only.
- Ready-to-use bi-plates are not to be considered a "sterile product" as they are not subject to terminal sterilization but a product with controlled bio contamination, within the limits of defined specifications reported on the Quality Control Certificate.
- Sterilize all biohazard waste before disposal. Dispose the unused medium and the plates inoculated with samples or microbial strains in accordance with current local legislation.
- The Certificates of Analysis and the Safety Data Sheet of the product are available on the website www.biolifeitaliana.it.
- The information provided in this document has been defined to the best of our knowledge and ability and represents a guideline for the proper use of the product but without obligation or liability. In all cases existing local laws, regulations and standard procedures must be observed for the examination of samples collected from human and animal organic districts, for environmental samples and for products intended for human or animal consumption. Our information does not relieve our customers from their responsibility for checking the suitability of our product for the intended purpose.

14 - STORAGE CONDITIONS AND SHELF LIFE

Upon receipt, store plates in their original pack at 2-8°C away from direct light. If properly stored, the plates may be used up to the expiration date. Do not use the plates beyond this date. Plates from opened plastic sachet can be used for 7 days when stored in a clean area at 2-8°C. Do not use the plates if the plastic sachet is damaged or if the dish is broken. Do not use the plates with signs of deterioration (e.g., microbial contamination, dehydration, shrinking or cracking of the medium, atypical colour, excess of moisture).

15 - REFERENCES

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TABLE OF APPLICABLE SYMBOLS

REF Catalogue	or REF e number	LOT	Batch code	IVD In vitro Diagnos Medical		Manufacturer	\square	Use by
1	Temperature limitation	\sum	Contents sufficient for <n> tests</n>	Consul Instruct Use	t ions for	For single use only		Fragile, handle with care

REVISION HISTORY

Version	Description of changes	Date	
Instructions for Use (IFU) - Revision 0	First edition, in compliance with IVDR 2017/746	2021/09	
Instructions for Use (IFU) - Revision 1	Removal of obsolete classifications	2023/03	

Note: minor typographical, grammatical, and formatting changes are not included in the revision history