Biolife

Instructions for use

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m-FAECAL COLIFORM AGAR

(m-FC Agar)

Dehydrated culture medium, ready to use plates

1 - INTENDED USE

m-Faecal Coliform Agar is used with rosolic acid for the cultivation and enumeration of faecal coliforms by the membrane filter technique.

2 - COMPOSITION - TYPICAL FORMULA * (AFTER RECONSTITUTION WITH 1 L OF WATER)

DEHYDRATED M-FAECAL COLIFORM AGAR			
Tryptose	10.0 g		
Peptocomplex	5.0 g		
Yeast extract	3.0 g		
Sodium chloride	5.0 g		
Lactose	12.5 g		
Bile salts N° 3	1.5 g		
Aniline blue	0.1 g		
Agar	13.0 g		
READY-TO-USE PLATES			
Tryptose	10.0 g		
Peptocomplex	5.0 g		
Yeast extract	3.0 g		
Sodium chloride	5.0 g		
Lactose	12.5 g		
Bile salts N. 3	1.5 g		
Aniline blue	0.1 g		
Agar	13.0 g		
Rosolic acid 1%	10 mL		
Purified water	1000 mL		

*The formula may be adjusted and/or supplemented to meet the required performances criteria.

3 - PRINCIPLE OF THE METHOD AND EXPLANATION OF THE PROCEDURE

Faecal coliforms are found in the gastro intestinal tracts and faeces of warm-blooded animals and may be differentiated from coliforms from environmental sources by their ability to grow at 44.5 °C.

m-Faecal Coliform Agar is prepared according to the formulation reported by Geldrich et al. in 1965¹ for the enumeration of faecal coliforms using the membrane filter technique without prior enrichment. The medium is recommended by many International Authorities for the enumeration of faecal coliforms using the membrane filter technique.²⁻⁴ Faecal coliforms (or thermotolerant coliform bacteria) are frequently used as indicators of faecal pollution although they are a less specific indicator of faecal contamination than *Escherichia coli*, since they may sometimes arise from nonfecal sources, especially in tropical climates.⁵

Tryptose and peptocomplex provide nitrogen and minerals for microbial growth, yeast extract is a source of B-vitamins complex for growth stimulation, lactose is a fermentable carbohydrate and a source of carbon and energy, sodium chloride maintains the osmotic balance. Bile salts N° 3 and aniline blue inhibit the growth of Gram-positive bacteria. The high incubation temperature makes the medium more selective. Aniline blue and rosolic acid form the indicator system of the medium.

4 - DIRECTIONS FOR DEHYDRATED MEDIUM PREPARATION

Suspend 50 g in 1000 mL of cold purified water. Heat to boiling with frequent agitation, and add 10 mL of Rosolic Acid (REF 4211901) 1% solution in NaOH 0.2 N and continue to boil for 1 minute. Do not sterilize in the autoclave. Cool to 47-50 °C, mix well and pour into sterile dishes for MF technique (55 mm diameter).

5 - PHYSICAL CHARACTERISTICS Dehvdrated medium appearan

Dehydrated medium appearance	yellowish, fine, homogeneous, free-flowing powder
Solution and prepared plates appearance	grey-violet, limpid
Final pH at 20-25 °C	7.4 ± 0.2

6 - MATERIALS PROVIDED - PACKAGING

Product	Туре	REF	Pack
m-Faecal Coliform Agar (m-FC Agar)	Dehydrated medium	4014872	500 g (10 L)
m-FC Agar	Ready-to-use plates	491487	3 x 10 plates ø 55 mm

7 - MATERIALS REQUIRED BUT NOT PROVIDED

Autoclave, water-bath, sterile loops and pipettes, incubator and laboratory equipment as required, Erlenmeyer flasks, sterile Petri dishes, membrane filters, Rosolic Acid (REF 4211901) ancillary culture media and reagents.

8 - SPECIMENS

Water samples. For sample collection, storage, transport and preparation, follow good laboratory practice and refer to applicable International Standards and regulations.

9 - TEST PROCEDURE

1. Filter an appropriate volume of water onto the membrane depending on the expected faecal coliforms number. When the sample's bacterial density is unknown, filter several volumes or dilutions to achieve a countable plate (20-60 CFU/dish).







2. Using aseptic technique, roll the membrane filter used to collect the water sample onto the surface of the agar, so as to avoid the formation of air bubbles between the filter and the agar surface.

3. Within 30 minutes place the dishes in plastic bags and incubate, by immersion, in a water bath at 44.5 ± 0.2°C for 24 ± 2 hours.

10 - READING AND INTERPRETATION

After incubation, observe bacterial growth and record any specific morphological and colour characteristic of the colonies. Count and record colonies with various shades of blue as faecal coliforms. Colonies of non-faecal coliforms are grey or cream or pink.

11 - USER QUALITY CONTROL

All manufactured lots of the product are released for sale after the Quality Control has been performed to check the compliance with the specifications. However, the end user can perform its own Quality Control in accordance with the local applicable regulations, in compliance with accreditation requirements and the experience of the Laboratory. Here below are listed some test strains useful for the quality control.

CONTROL STRAINS	INCUBATION T°/ T - ATM	EXPECTED RESULTS
E. coli ATCC 25922	44.5°/ 24 H-A	growth with blue colonies
E. faecalis ATCC 19433	44.5°/ 24 H-A	inhibited

A: aerobic incubation; ATCC is a trademark of American Type Culture Collection

12 – PERFORMANCES CHARACTERISTICS

Prior to release for sale, representative samples of all lots of dehydrated and ready-to-use plates of m-Faecal Coliform Agar (Test Batch:TB) are tested for productivity and selectivity by comparing the results with a previously approved Reference Batch (RB).

Productivity is tested by a quantitative method with the target strains *E. coli* ATCC 25922 and *E. coli* ATCC 8739 : the filters rolled on MF plates are inoculated with decimal dilutions in saline of a colonies' suspension and incubated at 44°C for 24 hours. The colonies are enumerated on both batches and the productivity ratio (Pr) is calculated. If Pr is \geq 0.7 and if the colonies morphology and colour are typical (blue colonies) the results are considered acceptable and conform to the specifications.

Selectivity and specificity are evaluated with modified Miles-Misra surface drop method by inoculating the plates with suitable decimal dilutions in saline of a 0.5 McFarland suspension of the following non-target strains: *S. aureus* ATCC 25923, *E,f aecalis* ATCC 19433, *K. pneumoniae* ATCC 27736, *C. freundii* ATCC 8090, *S.* Typhimurium ATCC 14024. The growth of Gram-positive strains is totally inhibited while the growth of *K. pneumoniae* and *C. freundii* is partially inhibited and the growth of *S*.Typhimurium is not inhibited and the strain grows with pink colonies.

13 – LIMITATIONS OF THE METHOD

- Since the incubation temperature is critical, the use of submerged waterproofed MF culture is recommended or the use of an incubator that is documented to hold the temperature at 44.5°C± 0.2°C throughout the chamber over a 24 hours period.²
- There are limitations to the interpretation of a thermotolerant coliform result from thermal waters and pulp and paper mill effluent samples where thermotolerant *Klebsiella* have predominated and not been indicative of a sewerage source. Approximately 60% to 80% of all *Klebisella* from faeces and clinical samples are positive in the thermotolerant coliform test and are *K. pneumoniae.*²

14 - PRECAUTIONS AND WARNINGS

- The medium is for microbiological control and for professional use only; it is to be used by adequately trained and qualified laboratory personnel, observing approved biohazard precautions and aseptic techniques.
- Dehydrated media must be handled with suitable protection. Before use, consult the Safety Data Sheet.
- This culture medium contains raw materials of animal origin. The ante and post mortem controls of the animals and those during the
 production and distribution cycle of the raw materials, cannot completely guarantee that this product doesn't contain any transmissible
 pathogen. Therefore, it is recommended that the culture medium be treated as potentially infectious, and handled observing the usual
 specific precautions: do not ingest, inhale, or allow to come into contact with skin, eyes, mucous membranes. Download the TSE
 Statement from the website www.biolifeitaliana.it, describing the measures implemented by Biolife Italiana for the risk reduction linked to
 infectious animal diseases.
- · Apply Good Manufacturing Practice in the production process of prepared media.
- Each plate of this culture medium is for single use only.
- Ready-to-use plates are not to be considered a "sterile product" as they are not subject to terminal sterilization, but a product with controlled bio contamination, within the limits of defined specifications reported on the Quality Control Certificate.
- All laboratory specimens should be considered infectious.
- The laboratory area must be controlled to avoid contaminants such as medium powder or microbial agents.
- Sterilize all biohazard waste before disposal. Dispose the unused medium and the sterilized medium inoculated with samples or microbial strains in accordance with current local legislation.
- Do not use the culture medium as active ingredient for pharmaceutical preparations or as production material intended for human and animal consumption.
- The Certificates of Analysis and the Safety Data Sheets are available on the website www.biolifeitaliana.it.
- The information provided in this document has been defined to the best of our knowledge and ability and represents a guideline for the
 proper use of the product but without obligation or liability. In all cases existing local laws, regulations and standard procedures must be
 observed for the examination of samples collected from human and animal organic districts, for environmental samples and for products
 intended for human or animal consumption. Our information does not relieve our customers from their responsibility for checking the
 suitability of our product for the intended purpose.

15 - STORAGE CONDITIONS AND SHELF LIFE

Dehydrated medium

Upon receipt, store at +10°C /+30°C away from direct light in a dry place. If properly stored, it may be used up to the expiration date. Do not use beyond this date. Avoid opening the bottle in humid places. After use, the container must be tightly closed. Discard the product if the container and/or the cap are damaged, or if the container is not well closed, or in case of evident deterioration of the powder (colour changes, hardening, large lumps).

The user is responsible for the manufacturing and quality control processes of prepared media and the validation of their shelf life, according to the type (plates/flasks) and the applied storage conditions (temperature and packaging).





Ready to use plates

Upon receipt, store plates in their original pack at $+2^{\circ}C / +8^{\circ}C$ away from direct light. If properly stored, the plates may be used up to the expiration date. Do not use the plates beyond this date. Plates from opened plastic sachet can be used for 7 days when stored in a clean area at $+2^{\circ}C / +8^{\circ}C$. Do not use the plates if the plastic sachet is damaged or if the dish is broken. Do not use the plates with signs of deterioration (e.g., microbial contamination, dehydration, shrinking or cracking of the medium, atypical colour, excess of moisture).

16 - REFERENCES

- 1. Geldreich EE, Clark HF, Huff CB, Best LC. Faecal coliform organisms medium for membrane filtration technique. J Am Water Works Assoc 1965; 57:208.
- APHA Standard methods for the examination of water and wastewater, 23rd ed. American Public Health Association, Washington, D.C., 2017.
 AOAC Official methods of analysis, 18th ed., AOAC International. Gaithersburg, Md. 2007
- U.S. Environmental Protection Agency. Manual for the certification of laboratories analyzing drinking water. EPA-814B-92-002. Office of Ground Water and Technical Support Division, USEPA, Cincinnati, Ohio.
- 5. Cisneros BJ, in Treatise on Water Science, 4.06.4.1.5 Biological indicators, 2011.

TABLE OF APPLICABLE SYMBOLS

REF or REF Catalogue number	LOT Batch code	Manufacturer	For single use only	☐ This side up	Store in a dry place
Temperature limitation	Content sufficient for <n> tests</n>	Consult Instruction s for Use	Use by	Fragile	Keep away from direct light

REVISION HISTORY

ſ	Version	Description of changes	Date			
Ē	Revision 1	Updated layout and content	01/2023			
No	Note: minor typographical, grammatical, and formatting changes are not included in the revision history.					

