

INSTRUCTIONS FOR USE

SABOURAUD DEXTROSE AGAR CAF 500

Dehydrated culture medium



1 - INTENDED USE

In vitro diagnostic. Selective medium for the isolation of yeasts and moulds in clinical specimens.

2 - COMPOSITION - TYPICAL FORMULA *

(AFTER RECONSTITUTION WITH 1	L OF WATER)
Pancreatic digest of casein	5.0 g
Peptic digest of meat	5.0 g
Glucose	40.0 g
Agar	15.0 g
Chloramphenicol	0.5 g

*The formula may be adjusted and/or supplemented to meet the required performances criteria.

Sabouraud Dextrose Agar CAF 500: Aspergillus sp.

3 - PRINCIPLE OF THE METHOD AND EXPLANATION OF THE PROCEDURE

By the end of the 1890's, Raymond Jacques Sabouraud had crystallized and organized the scattered observations regarding the role of pathogenic fungi in dermatophytic infections and proposed a medium for their isolation and classification.^{1,2}

Numerous experiments were performed to improve Sabouraud's formula with a variety of peptones and carbohydrates by Weidman and Spring³, but the suitable medium was selected by Hodges⁴. In its final formulation, this medium contained 1% peptone, 4% dextrose, and 1.8% agar, with a final pH of 5.0. This formulation was named Sabouraud medium and is the basic routine culture medium used to grow fungi in clinical laboratories.

The components of Sabouraud Dextrose Agar conform to the recommendations of the current European Pharmacopoeia⁵. The addition of chloramphenicol at the concentration of 500 mg/L is a modification designed to increase bacterial inhibition and improve the isolation of opportunistic fungi from contaminated specimens.

Sabouraud Dextrose Agar CAF 500 is a selective medium for the isolation of yeasts and moulds, mainly opportunist pathogens (*Aspergillus, Fusarium, Mucor, Rhizopus*, etc.), cycloheximide sensitive fungi such as *Cryptococcus neoformans* and *Allescheria boydii* and *Candida* spp. in clinical specimens.

Pancreatic digest of casein and peptic digest of meat provide nitrogen, carbon and trace elements for microbial growth. Glucose, at high concentration is a carbon and energy source. The low pH is favourable for the growth of fungi and is slightly inhibitory to contaminating bacteria; the selective properties are increased by the presence of chloramphenicol, a broad-spectrum antibiotic, which is inhibitory to a wide range of Gram-negative and Gram-positive bacteria. The higher concentration of chloramphenicol (500 mg/L) compared to standard formulations (50 or 100 mg/L) is not inhibitory of fungal growth and increases the selective properties of the medium towards bacterial contaminants, especially *Pseudomonas*.⁶

4 - DIRECTIONS FOR MEDIUM PREPARATION

Suspend 65.5 g in 1000 mL of cold purified water. Heat to boiling with frequent agitation and sterilize by autoclaving at 121°C for 15 minutes. Cool to 47-50°C, mix well and pour into sterile Petri dishes. Notes

Do not exceed the boiling and sterilization times and temperatures.

Alternatively distribute in screw capped tubes before sterilization and solidify in slanted position.

5 - PHYSICAL CHARACTERISTICS

Dehydrated medium appearance	yellow, fine, homogeneous, free-flowing powder
Solution and prepared plates appearance	yellow, limpid
Final pH at 20-25 °C	5.6 ± 0.2

6 - MATERIALS PROVIDED

UNATERIAEOT ROVIDED			
Product	Туре	REF	Pack
Sabouraud Dextrose Agar CAF 500	Dehydrated medium	4020072	500 g (7,6 L)

7 - MATERIALS REQUIRED BUT NOT PROVIDED

Autoclave, water-bath, sterile loops and swabs, incubator and laboratory equipment as required, Petri dishes, Erlenmeyer flasks, ancillary culture media and reagents for the identification of the colonies.

8 - SPECIMENS

Sabouraud Dextrose Agar CAF 500 can be directly inoculated with many clinical specimens collected from various normally non-sterile human sites. Refer to the quoted literature for specimen types, related to specific infections.⁷⁻⁸ Sabouraud Dextrose Agar CAF 500 is not suitable for direct inoculation of blood samples or other specimens from normally sterile sites. Collect specimens before antimicrobial therapy where possible. Good laboratory practices for collection, transport and storage of the clinical specimens should be applied; consult appropriate references for further information.⁷





9 - TEST PROCEDURE

Allow plates to come to room temperature and to dry the surface of the medium. Inoculate the clinical specimen as soon as possible after collection; streak with a loop over the four quadrants of the plate to obtain well isolated colonies. Alternatively, if the material is being cultured directly from a swab, roll the swab over a small area of the surface at the edge; then streak from this inoculated area. For cutaneous samples, press specimen lightly into medium.

Inoculate each specimen in duplicate; incubate one set in aerobic condition at 22-25°C, the other at 33-37°C.9

For dermatophytes, examine cultures every 4-6 days for a period of up to 20 days; for others incubate 2-5 days. Plates should be incubated under conditions of increased humidity during prolonged incubation.

The user is responsible for choosing the appropriate incubation time and temperature depending on the processed specimen, the requirements of organisms to be recovered and the local applicable protocols.

10 - READING AND INTERPRETATION

Clinical specimen: after incubation, observe the bacterial growth and record the specific morphological and chromatic characteristics of the colonies and sub-culture to appropriate media for further identification tests.

11 - USER QUALITY CONTROL

All manufactured lots of the product are released for sale after the Quality Control has been performed to check the compliance with the specifications. However, the end user can perform its own Quality Control in accordance with the local applicable regulations, in compliance with accreditation requirements and the experience of the Laboratory. Here below are listed some test strains useful for the quality control.¹⁰

T.mentagrophytes ATCC 9533 $20-25^{\circ}C / \le 5$ days/ Agood growth, white colonies with typical morphologyA.brasiliensis ATCC 16404 $20-25^{\circ}C / \le 5$ days/ Agood growth, white/black colonies with typical morpholE.coli ATCC 25922 $20-25^{\circ}C / \le 5$ days/ Ainhibited	CONTROL STRAINS	INCUBATION $T^{\circ}/T / ATM$	EXPECTED RESULTS
	C.albicans ATCC 10231	20-25°C / \leq 5 days/ A	good growth, white yeast-like colonies
	T.mentagrophytes ATCC 9533	20-25°C / \leq 5 days/ A	good growth, white colonies with typical morphology
	A.brasiliensis ATCC 16404	20-25°C / \leq 5 days/ A	good growth, white/black colonies with typical morpholog
	E.coli ATCC 25922	20-25°C / \leq 5 days/ A	inhibited

A: aerobic incubation; ATCC is a trademark of American Type Culture Collection

12 - PERFORMANCES CHARACTERISTICS

Prior to release for sale a representative sample of all lots of dehydrated Sabouraud Dextrose Agar CAF 500 is tested for productivity and selectivity by comparing the results with a previously approved Reference Batch.

Productivity is tested by semi-quantitative ecometric technique with the target strains *C.albicans* ATCC 10231, *A.brasiliensis* ATCC 16404, *S.cerevisiae* ATCC 9763, *P.chrysogenum* ATCC 10106, *T.mentagrophytes* ATCC 9533, *M.canis* ATCC 36299.

After incubation at 20-25°C for up to 3 days, the amount of growth on the plates and colonies' characteristics are evaluated and recorded: they shall be comparable in both batches.

The selectivity is evaluated with modified Miles-Misra surface drop method by inoculating the plates with suitable decimal dilutions in saline of a 0.5 McFarland suspension of the non-target strains *E.coli* ATCC 25922, *P.aeruginosa* ATCC 14207 and *S.aureus* ATCC 25923. The growth of non-target strains is totally inhibited.

13 - LIMITATIONS OF THE METHOD

- Chloramphenicol may inhibit some pathogenic fungi (e.g, Actinomyces bovis and Nocardia asteroids).^{6,9}
- Sabouraud Dextrose Agar CAF 500 has a poor efficacy in the isolation of Histoplasma capsulatum from potentially contaminated clinical specimens.¹¹
- A single medium is only rarely useful to recover all pathogens contained in a specimen. Therefore, additional media for the isolation of yeasts and moulds with lower selectivity such as Sabouraud Dextrose Agar or Potato Dextrose Agar and with different selective compounds such as Dermatophyte Test medium, should be used.
- Even if the microbial colonies on the plates are differentiated on the basis of their morphological and chromatic characteristics, it is recommended that biochemical, immunological, molecular, or mass spectrometry testing be performed on isolates, from pure culture, for complete identification. If relevant, perform antimicrobial susceptibility testing.
- This culture medium is intended as an aid in the diagnosis of infectious diseases; the interpretation of the results must be made considering the patient's clinical history, the origin of the sample and the results of other diagnostic tests.

14 - PRECAUTIONS AND WARNINGS

- This product is a qualitative *in vitro* diagnostic, for professional use only; it is to be used by adequately trained and qualified laboratory personnel, observing approved biohazard precautions and aseptic techniques.
- Dehydrated media must be handled with suitable protection. Before use, consult the Safety Data Sheet.
- This culture medium contains raw materials of animal origin. The ante and post mortem controls of the animals and those during the production and distribution cycle of the raw materials, cannot completely guarantee that this product doesn't contain any transmissible pathogen. Therefore, it is recommended that the culture medium be treated as potentially infectious, and handled observing the usual specific precautions: do not ingest, inhale, or allow to come into contact with skin, eyes, mucous membranes. Download the TSE Statement from the website www.biolifeitaliana.it, describing the measures implemented by Biolife Italiana for the risk reduction linked to infectious animal diseases.
- Apply Good Manufacturing Practice in the preparation process of plated or tubed or bottled media.
- All laboratory specimens should be considered infectious.
- The laboratory area must be controlled to avoid contaminants such as culture medium or microbial agents.
- Sterilize all biohazard waste before disposal. Dispose the unused medium and the sterilized plates inoculated with samples or microbial strains in accordance with current local legislation.
- Do not use the culture medium as active ingredient for pharmaceutical preparations or as production material intended for human and animal consumption.
- The Certificates of Analysis and the Safety Data Sheet of the product are available on the website www.biolifeitaliana.it.
- Notify Biolife Italiana Srl (complaint@biolifeitaliana.it) and the relevant Authorities of any serious incident occurring in connection with the use of the *in vitro* diagnostic.





The information provided in this document has been defined to the best of our knowledge and ability and represents a guideline for the
proper use of the product but without obligation or liability. In all cases existing local laws, regulations and standard procedures must be
observed for the examination of samples collected from human and animal organic districts, for environmental samples and for products
intended for human or animal consumption. Our information does not relieve our customers from their responsibility for checking the
suitability of our product for the intended purpose.

15 - STORAGE CONDITIONS AND SHELF LIFE

Upon receipt, store at +10°C /+30°C away from direct light in a dry place. If properly stored, it may be used up to the expiration date. Do not use beyond this date. Avoid opening the bottle in humid places. After use, the container must be tightly closed. Discard the product if the container and/or the cap are damaged, or if the container is not well closed, or in case of evident deterioration of the powder (colour changes, hardening, large lumps). The user is responsible for the manufacturing and quality control processes of prepared media and for the validation of the shelf life of the finished products, according to the type (plates/tubes/bottles), and the storage method applied (temperature and packaging).

16 - REFERENCES

- 1. Espinel-Ingroff A. History of medical mycology in the United States. Clin Microbiol Rev 1966;9:235-272
- Sabouraud R. Contribution à l'étude de la trichophytie humaine. Etude clinique, microscopique et bactériologique sur la pluralité des trichophytons de l'homme. Ann Dermatol Syphil 1892; 3:1061-1087.
- 3. Weidman FD, Spring D. Comparison of ringworm culture ingredients: II and III. Arch Dermatol Syphilol 1928; 18:829–851.
- 4. Hodges RS. Cultures of ringworm fungi on Sabouraud's proof mediums and on mediums prepared with American peptones and sugars. Arch Dermatol Syphilol 1928;18:852–856.
- 5. European Pharmacopoeia, current edition
- McLean Jr IW, Schwab JL, Hillegas AB, Schlingman AS. Susceptibility of microorganisms to chloramphenicol (chloromycetin). J Clin Invest. 1949; 28(5):953-963.
- Berkow EL, McGowan KL. Specimen Collection, Transport and Processing: Mycology. In Carrol KC, Pfaller MA et al. editors. Manual of clinical microbiology, 12th ed. Washington, DC: American Society for Microbiology; 2019.
- 8. Public Health England- UK SMI B 17: tissues and biopsies from deep-seated sites and organs. 05.01.18
- 9. MacFaddin JF. Media for Isolation-Cultivation-Identification-Maintenance of Medical Bacteria. Baltimore: Williams & Wilkins; 1985.
- 10. CLSI (formerly NCCLS) Quality Control of Commercially Prepared Culture Media. Approved Standard, 3rd edition. M22 A3 vol. 24 n° 19, 2004
- 11. Unis G, da Silva VB, Severo LC. Disseminated histoplasmosis and AIDS. The role of culture medium for the bronchoscopic clinical specimens Rev Soc Bras Med Trop. 2004;37:234-7

TABLE OF APPLICABLE SYMBOLS



REVISION HISTORY

Version	Description of changes	Date
Revision 1	Updated layout and content	2022/04
Revision 2	Removal of obsolete classification	2023/04

Note: minor typographical, grammatical, and formatting changes are not included in the revision history.

