



## ENDO BROTH MEMBRANE FILTER (m-ENDO BROTH) Dehydrated culture medium

### 1 - INTENDED USE

For the enumeration of coliforms in water samples by membrane filtration.

### 2 – COMPOSITION\*

#### TYPICAL FORMULA (AFTER RECONSTITUTION WITH 1 L OF WATER)

Yeast extract	1.500 g
Tryptone	5.000 g
Peptone	5.000 g
Tryptose	10.000 g
Lactose	12.500 g
Dipotassium hydrogen phosphate	4.375 g
Potassium dihydrogen phosphate	1.375 g
Sodium chloride	5.000 g
Sodium deoxycholate	0.100 g
Sodium lauryl sulphate	0.050 g
Sodium sulphite	2.100 g
Basic fuchsin	1.050 g

\*The formula may be adjusted and/or supplemented to meet the required performances criteria.

### 3 - PRINCIPLE OF THE METHOD AND EXPLANATION OF THE PROCEDURE

Endo media were originally developed by Endo<sup>1</sup> for the isolation of the typhoid bacillus. McCarthy, Delaney, and Grasso<sup>2</sup> modified Endo's formulation and proposed the LES (Lawrence Experimental Station) Endo media, for the recovery of coliforms with a membrane filter two steps technique.

Both one and two steps membrane filter procedures have been included in the APHA Standard Methods for the detection of coliforms in drinking, non-potable, and other waters.<sup>3</sup>

Endo Broth Membrane Filter (m-Endo Broth) has a formulation with the same components as LES Endo Agar, at slightly different concentrations and without agar.

In m-Endo Broth, essential growth factors are provided by peptones which are sources of nitrogen, carbon and minerals. Yeast extract is a source of vitamins, particularly of the B-group. Phosphates are used as buffering agents to control the pH in the medium. Sodium chloride is a source of electrolytes and maintains the osmotic equilibrium. The slight inhibition of Gram-positive bacteria achieved with the sodium sulphite/acid fuchsin combination in classical Endo formulation, has been improved in "Endo Broth" formulation by inclusion of sodium deoxycholate and sodium lauryl sulphite. The sodium sulphite in the medium also has the function of decolourising acid fuchsin as it does in Schiff's reagent. Lactose-fermenting bacteria produce acetaldehyde from lactose which releases the fuchsin from the colourless fuchsin-sulphite compound and colours the colonies red; when the reaction is rapid and very intense (e.g in the case of *E. coli*), the fuchsin crystallises and produces a metallic sheen on the colonies. In areas of the plate with intense growth, the metallic sheen is suppressed. Non-lactose-fermenting organisms produce colourless colonies against the pink background of the medium.

### 4 - DIRECTIONS FOR DEHYDRATED MEDIUM

Suspend 48 g in 1000 mL of cold purified water. Add 20 mL of 95% ethanol, heat to boiling with frequent agitation to dissolve completely. Do not autoclave, do not overboil. Cool to 47-50°C, mix well for resuspending the precipitate.

### 5 - PHYSICAL CHARACTERISTICS

Dehydrated medium appearance	purple, fine, homogeneous, free-flowing powder with small dark particles
Solution appearance	pink-orange, slightly opalescent to opalescent with small dark particles
Final pH at 20-25 °C	7.2 ± 0.2

### 6 - MATERIALS PROVIDED - PACKAGING

Product	Type	REF	Pack
m- Endo Broth	Dehydrated medium	4014612	500 g (10.41 L)

### 7 - MATERIALS REQUIRED BUT NOT PROVIDED

Water-bath, sterile loops and pipettes, incubator and laboratory equipment as required, Erlenmeyer flasks, sterile Petri dishes, 95% ethanol, membrane filtration system, ancillary culture media and reagents.

### 8 – SPECIMENS

Water samples. Consult the appropriate references for sample collection, storage and preparation.<sup>3</sup>

### 9 - TEST PROCEDURE

#### One step technique<sup>3</sup>

1. Place an absorbent pad in a 55 mm Petri dish and pipette at least 2-3 mL of m-Endo Broth to saturate pad.
2. Using an appropriate sterile filtration unit, filter the water sample.
3. Aseptically, place the membrane filter on the pad avoiding the formation of air bubbles between the filter and the pad surface.
4. Incubate at 35°C for 22 to 24 hours.

#### Two steps technique<sup>3</sup>

1. Place an absorbent pad in a 55 mm Petri dish and pipette at least 2 mL of Lauryl Pepto Bios Broth (REF 401580), to saturate pad.
2. Using an appropriate sterile filtration unit, filter the water sample aseptically, place the membrane filter on the pad and incubate for 1.5 - 2 hours at 35°C in a moist atmosphere.





3. Transfer the membrane from the pad to 55 mm Petri dish containing a pad saturated with at least 2-3 mL of m-Endo Broth, avoiding the formation of air bubbles between the filter and the agar surface. Incubate at 35°C for 22 to 24 hours.

#### 10 - READING AND INTERPRETATION

After incubation, observe the bacterial growth and record the specific morphological and chromatic characteristics of the colonies.

Typical coliforms colonies are pink to red with metallic sheen. The sheen may cover the entire colony or may only appear in the centre or on the periphery.

Some colonies will appear pink or red but lack the characteristic metallic sheen. These colonies are classified as atypical coliforms and need to be verified through further testing.

Typical non-lactose fermenters colonies are colourless against the pink-red background of the medium

#### 11 - USER QUALITY CONTROL

All manufactured lots of the product are released for sale after the Quality Control has been performed to check the compliance with the specifications. However, the end user can perform its own Quality Control in accordance with the local applicable regulations, in compliance with accreditation requirements and the experience of the Laboratory. Here below are listed some test strains useful for the quality control.

CONTROL STRAINS	INCUBATION T° / T / ATM	EXPECTED RESULTS
<i>E. coli</i> ATCC 25922	37°C/24H-A	good growth, pink-red colonies with metallic sheen
<i>S. Enteritidis</i> ATCC 13076	37°C/24H-A	good growth, colourless colonies

A: aerobic incubation; ATCC is a trademark of American Type Culture Collection

#### 12 – PERFORMANCE CHARACTERISTICS

Prior to release for sale, a representative sample of all lots of dehydrated m-Endo Broth (Test Batch TB) is tested for productivity, specificity and selectivity by comparing the results with a previously approved Reference Batch (RB).

The productivity is tested by a quantitative method with the target strains *E. coli* ATCC 25922 and *E. aerogenes* ATCC 13048: the membrane filters on the plates are inoculated with decimal dilutions in saline of a colonies' suspension and incubated at 37°C for 24 hours. The colonies are enumerated on both batches and the productivity ratio ( $Pr:CFU_{TB}/CFU_{RB}$ ) is calculated. If  $Pr \geq 0.7$  and if the colonies morphology and colour are typical (pink-red colonies with metallic sheen) the results are considered acceptable and conform to the specifications.

Moreover, the productivity and specificity characteristics are tested by semi-quantitative ecometric technique with the following strains: *E. coli* ATCC 8739, *K. pneumoniae* ATCC 23357 and *S. Enteritidis* ATCC 13086. After incubation, the amount of growth and the colony characteristics are evaluated: coliforms strains exhibit good growth with pink-red colonies with metallic sheen while *S. Enteritidis* grows with colourless colonies.

The selectivity is assessed by modified Miles-Misra surface drop method by inoculating the plates with suitable decimal dilutions in saline of a 0.5 McFarland suspension of *S. aureus* ATCC 25923 and *E. faecalis* ATCC 19433. The growth of Gram-positive strains is totally inhibited.

#### 13-LIMITATIONS OF THE METHODS

- Occasionally, non-coliform organisms may produce typical shining colonies.
- Occasionally, some colonies will appear pink or red but lack the characteristic metallic sheen. These colonies are classified as atypical coliforms and need to be verified with further tests.

#### 14 - PRECAUTIONS AND WARNINGS

- This culture medium is for microbiological control and for professional use only; it is to be used by adequately trained and qualified laboratory personnel, observing approved biohazard precautions and aseptic techniques.
- Dehydrated m-Endo Broth is classified as dangerous since contains acid fuchsin, a potential carcinogen. Before use, consult the Safety Data Sheet.
- This culture medium contains raw materials of animal origin. The *ante* and *post mortem* controls of the animals and those during the production and distribution cycle of the raw materials, cannot completely guarantee that this product doesn't contain any transmissible pathogen. Therefore, it is recommended that the culture medium be treated as potentially infectious, and handled observing the usual specific precautions: do not ingest, inhale, or allow to come into contact with skin, eyes, mucous membranes. Download the TSE Statement from the website [www.biolifeitaliana.it](http://www.biolifeitaliana.it), describing the measures implemented by Biolife Italiana for the risk reduction linked to infectious animal diseases.
- Apply Good Manufacturing Practice in the production process of prepared media.
- All laboratory specimens should be considered infectious.
- The laboratory area must be controlled to avoid contaminants such as medium powder or microbial agents.
- Sterilize all biohazard waste before disposal. Dispose the unused medium and the sterilised inoculated medium with samples or microbial strains in accordance with current local legislation.
- Do not use the culture medium as active ingredient for pharmaceutical preparations or as production material intended for human and animal consumption
- The Certificates of Analysis and the Safety Data Sheet of the products are available on the website [www.biolifeitaliana.it](http://www.biolifeitaliana.it).
- The information provided in this document has been defined to the best of our knowledge and ability and represents a guideline for the proper use of the product but without obligation or liability. In all cases existing local laws, regulations and standard procedures must be observed for the examination of samples collected from human and animal organic districts, for environmental samples and for products intended for human or animal consumption. Our information does not relieve our customers from their responsibility for checking the suitability of our product for the intended purpose.

#### 15 - STORAGE CONDITIONS AND SHELF LIFE

Upon receipt, store at +10°C /+30°C away from direct light in a dry place. If properly stored, it may be used up to the expiration date. Do not use beyond this date. Avoid opening the bottle in humid places. After use, the container must be tightly closed. Discard the product if the container and/or the cap are damaged, or if the container is not well closed, or in case of evident deterioration of the powder (colour changes, hardening, large lumps).

The user is responsible for the manufacturing and quality control processes of prepared media and the validation of their shelf life, according to the type and the applied storage conditions (temperature and packaging). According to APHA, the bottled medium may be stored at +2-+8°C in the dark, for up to 96 hours.<sup>3</sup>












### 16 - REFERENCES

1. Endo S. Über ein Verfahren zum Nachweis der Typhus bacillen. Centr f Bakt 1904; 35:109-110.
2. McCarthy JA, Delaney JE, Grasso RJ. Measuring coliforms in water. Water Sewage Works 1961; 108:238
3. APHA. Standard methods for the examination of water and wastewater, 23st ed., 2017. American Public Health Association, Washington, D.C.

### TABLE OF APPLICABLE SYMBOLS

<b>REF</b> or REF Catalogue number	<b>LOT</b> Batch code	 Manufacturer	 Store in a dry place	 Use by
 Temperature limitation	 Contents sufficient for <n> tests	 Consult Instructions for Use	 Keep away from direct light	

### REVISION HISTORY

Version	Description of changes	Date
Revision 1	Updated layout and content	2022/08

Note: minor typographical, grammatical, and formatting changes are not included in the revision history.

